Mussel Coating Protein-Derived Complex Coacervates Mitigate Frictional Surface Damage

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ABSTRACT: The role of friction in the functional performance of biomaterial interfaces is widely reckoned to be critical and complicated but poorly understood. To better understand friction forces, we investigated the natural adaptation of the holdfast or byssus of mussels that live in high-energy surf habitats. As the outermost covering of the byssus, the cuticle deserves particular attention for its adaptations to frictional wear under shear. In this study, we coacervated one of three variants of a key cuticular component, mussel foot protein 1, mfp-1 [(1) Mytilus californianus mcfp-1, (2) rmfp-1, and (3) rmfp-1-Dopa], with hyaluronic acid (HA) and investigated the wear protection capabilities of these coacervates to surfaces (mica) during shear. Native mcfp-1/HA coacervates had an intermediate coefficient of friction (µ > 0.3) but conferred excellent wear protection to mica with no damage from applied loads, Fp, as high as 300 mN (pressure, Pp > 2 MPa). Recombinant rmfp-1/HA coacervates exhibited a comparable coefficient of friction (µ > 0.3); however, wear protection was significantly inferior (damage at Fp > 60 mN) compared with that of native protein coacervates. Wear protection of rmfp-1/HA coacervates increased 5-fold upon addition of the surface adhesive group 3,4-dihydroxyphenylalanine, (Dopa). We propose a Dopa-dependent wear protection mechanism to explain the differences in wear protection between coacervates. Our results reveal a significant untapped potential for coacervates in applications that require adhesion, lubrication, and wear protection. These applications include artificial joints, contact lenses, dental sealants, and hair and skin conditioners.

KEYWORDS: biomimetic, adhesion, wear protection, interface, Mytilus californianus foot protein 1, mcfp-1, hyaluronic acid, HA

1. INTRODUCTION

California mussels (Mytilus californianus) live sedentary lives along wind- and wave-swept shores where waves (with velocities up to 25 m/sec) impose unrelenting lift and drag forces that threaten mussel dislodgement. Mussels resist dislodgement via an adaptive, fibrous holdfast known as the byssus (Figure 1a). Byssal threads mitigate wave impact by dissipating up to 72% of the applied wave energy, thereby mediating sessile attachment. Friction is key to mediating energy dissipation; yet understanding the underlying phenomena relating to energy dissipation in friction is a long-standing and challenging issue in physical science. Given the vast applications of friction, lubrication, and wear protection, there is much interest in understanding and implementing relevant design concepts from nature. California mussels are an excellent model system for understanding adaptive mechanisms of energy dissipation and friction under shear. A deeper understanding of the mussel byssal attachment system continues to inspire new materials including adhesives, coatings, and hydrogels.

Although the majority of the literature focuses on adhesive forces, frictional forces arguably contribute more to holdfast performance. For example, when a single attached byssal thread with a plaque is pulled normal to the surface (e.g., lift), detachment resistance is largely adhesive, whereas when pulled at low angles to the surface (e.g., drag), detachment resistance is an order of magnitude greater due to the added contribution of friction. Mussels have adaptations at multiple length scales (nm–cm) that utilize friction to dissipate energy. These adaptations include, but are not limited to, the radial distribution of anchoring threads, the spatulate morphology of plaques, the stiff to compliant thread gradient, the reversible plastic deformation of threads in tension, and an outer protective coating called the cuticle composed of hard spherical granules (diam 0.2 to 1 μm) dispersed in a continuous compliant matrix. Frictional forces, however, are highly context-dependent and can also lead to stress concentration, damage, and catastrophic failure.

As the outermost covering of the byssus, the cuticle deserves particular attention for its adaptations against three types of...
frictional damage: (1) abrasion of the exterior surface by sand, (2) interface deformation between the harder cuticle ($E_i = 2$ GPa) and the softer collagenous core ($E_i = 0.4$ GPa), and (3) deformation of the matrix sliding between the granules. The cuticle is remarkable for its ability to avoid catastrophic failure through microcracking and reversible deformation. In *M. californianus*, the cuticle is stiff ($E_i = 2$ GPa) yet extensible (120%), making it one of the most energy-tolerant materials known.

From a molecular perspective, the cuticle granules are preformed by condensation of mussel foot protein-1 (mcfp-1) through extensive Dopa-Fe$^{3+}$-complexation in the accessory gland. mcfp-1 is a highly modified, positively charged, intrinsically unstructured protein with 72 decapeptide repeats of the consensus sequence PKISYP**P**TY**K where P** is hydroxyproline, P** is dihydroxyproline, and Y** is dihydroxyphenylalanine (Dopa). Upon material secretion, the cuticle granules are coated by the matrix, producing the dispersion evident in Figure 1b. The matrix consists of mcfp-1 and an acidic, chymotrypsin-labile, Ca$^{2+}$-binding protein.

These two polyelectrolytes in the matrix are thought to be delivered as a complex coacervate.

Coacervation is an emerging theme in protein processing as used by mussels, sandcastle worms, and caddisfly larvae to concentrate, separate, and deposit polyelectrolytes underwater. Complex coacervation relies on Coulombically driven interactions between oppositely charged groups on distinct polyelectrolytes (polycations and polyanions) to bring about phase separation, whereby the polyelectrolytes become concentrated in a dense phase that settles on and coats surfaces underwater. In the mussel byssus, coacervates are proposed to both deliver adhesive proteins (mfp-3s) and form the cuticle matrix.

As a tough and energy-dissipative material, the cuticle provides an excellent model of frictional wear protection. We therefore created cuticle matrix mimics and used the surface forces apparatus (SFA) to investigate friction and surface damage mitigation. We made matrix mimics via complex coacervation of mcfp-1 (or its recombinant homologue) and hyaluronic acid (HA), a readily available biopolymer that is acidic, binds calcium, and has a mass that is well matched to that of mfp-1. Coacervated mcfp-1/HA from *M. californianus* robustly mitigated surface damage to mica, far surpassing (>4 fold) that of coacervated recombinant homologues and native mcfp-1/HA coacervates at uneven conditions.
mixing ratios. During compressive shear, surface damage mitigation was independent of the coefficient of friction but instead depended critically on surface adhesive groups (Dopa) and coacervation conditions. We propose that a surface adhesive layer protects the surface from damage by shifting the shear slip-plane away from the surface and into the bulk. This exceptional damage mitigation expands the role of complex coacervates in mussel byssus from fluidic intermediates in material processing to a functional structure with energy-dissipative and protective properties.

2. RESULTS

2.1. Complex Coacervation of Native mcfp-1 and rmfp-1 with HA. We measured coacervate formation via solution turbidity and transmittance at a wavelength of 600 nm.
varying mixing ratios peak at a 1:1 weight-to-weight mixing ratio, which corresponds to a 0.71:1 polycation-to-polyanion charge ratio for both mcfp-1/HA and rmfp-1/HA coacervates (Figure 2b and Materials and Methods, section 5.4). We performed rheology and tribology measurements at an ionic strength with maximum turbidity (10 mM for rmfp-1/HA coacervates and 60 mM for mcfp-1/HA coacervates) and at both optimal (1:1) mixing ratios and suboptimal (7:3, and 3:7) mixing ratios with respect to coacervate yield.

Turbidity measures a metastable (time dependent) process. Upon mixing the polycation and polyanion, the turbidity increased due to microdroplet formation (time = t = 0–0.25 min), after which turbidity was metastable (t = 0.25–20 min). Finally, turbidity decreased as microdroplets coalesced with other microdroplets, settled on, and coated the cuvette surface (t = 20 min–2 d). Therefore, we measured turbidity 1 min post-mixing, well within this metastable regime.

While transmittance measurements are unable to distinguish precipitates from coacervates, we confirmed a liquid coacervate phase by observing spherical, dispersed microdroplets (Figure 3).26 Furthermore, we confirmed a single, continuous coacervate phase by observing a uniform refractive index between the two mica surfaces, creating smooth Fringes of Equal Chromatic Order (FECO) in the Surface Forces Apparatus (SFA) (Figure 4).24

2.2. Energy Dissipation, \( E_D \), and Interfacial Energy, \( \gamma_{eff} \), of mcfp-1/HA and rmfp-1/HA Coacervates. We measured normal forces, \( F_D \), at varying distances, D, between two mica surfaces in the SFA with physisorbed coacervates in the presence of bulk solution (schematic shown in Figure 4a). We then calculated the adhesive/cohesion forces, \( F_{ad} \), using the equation \( F_{ad} = F_D/R \) where R is the radius of surface curvature. Smooth, continuous pull-outs in the \( F_{ad} \) vs D plots imply dissipative-bridging interactions. Integration of these pull-out curves quantifies the dissipative energy, \( E_D \).27 We only observed dissipative-bridging interactions in coacervates containing Dopa. mcfp-1/HA coacervates demonstrated dissipative-bridging interactions at all mixing ratios (mcfp-1/HA 1:1 ratio, \( E_D = 32 \pm 15 \text{ mJ/m}^2 \); 7:3 ratio, \( 58 \pm 53 \text{ mJ/m}^2 \); 3:7 ratio, \( 42 \pm 12 \text{ mJ/m}^2 \), Figure 5a). We observed dissipative-bridging interactions (\( E_D = 63 \pm 18 \text{ mJ/m}^2 \)) with rmfp-1-Dopa/HA coacervates, whereas sharp jump-outs (no dissipative-bridging interactions) were observed between surfaces with rmfp-1/HA coacervates lacking Dopa (Figure 5b).

The ability of a coacervate to coat a surface depends critically on its adhesion/cohesion energy, \( E_{ad} \), and the interfacial energy, \( \gamma_{eff} \). We calculated the interfacial energy using the minimum \( P_{ad} \) (maximum force required to separate the two surfaces) and the equation \( \gamma_{eff} = F_{ad} / 3dR \).28 The interfacial energy of all coacervates tested was low (7:3 mcfp-1/HA; \( \gamma_{eff} = 0.61 \pm 0.41 \text{ mJ/m}^2 \), 3:7 mcfp-1/HA; \( \gamma_{eff} = 0.43 \pm 0.37 \text{ mJ/m}^2 \), rmfp-1/HA; \( \gamma_{eff} = 0.34 \pm 0.11 \text{ mJ/m}^2 \), and rmfp-1-Dopa/HA; \( \gamma_{eff} = 0.72 \pm 0.17 \text{ mJ/m}^2 \) (Figure 5a and b)). Moreover, mcfp-1/HA coacervates at a 1:1 mixing ratio had the lowest interfacial energy (\( \gamma_{eff} = 0.28 \pm 0.08 \text{ mJ/m}^2 \)) of all the coacervates tested.

2.3. Tribology and Rheology of mcfp-1/HA and rmfp-1/HA Coacervates. We used the SFA to measure the lateral forces, \( F_D \), to determine the tribological and rheological properties of our coacervates. Here, wear protection is as the maximum load, \( F_D \), that coacervate-coated mica surfaces could withstand before splitting of the FECO idicated shear-induced damage to the mica surface (Figure 4b). Neither mcfp-1 nor HA in uncoacervated solution protected the surfaces from damage any more than the buffer alone (surfaces were damaged \( \sim 10 \text{ mN} \)).25,26 mcfp-1/HA (1:1) coacervates protected mica from damage, even at the maximum applied load, \( F_D \) of 300 mN (corresponding to a pressure \( >2 \text{ MPa} \), Figure 6). mcfp-1/HA coacervates with uneven mixing ratios failed to protect the mica surface at 4-fold lower loads (7:3 ratio, \( F_D = 73 \pm 53 \text{ mN} \); 3:7 ratio, \( F_D = 53 \pm 18 \text{ mN} \), rmfp-1-Dopa/HA protected surfaces from wear under 5-fold higher loads (\( F_D = 57 \pm 18 \text{ mN} \)) than recombinant coacervates without Dopa (\( F_D = 11 \pm 3 \text{ mN} \), Figure 6). All coacervate systems exhibited a similar coefficient of friction (\( \mu \sim 0.3 \), Figure 6, inset) during shear, save one: 7:3 mcfp-1/HA (\( \mu = 0.5 \pm 0.07 \)). To investigate reversibility, we increased the normal force which increased the lateral force, \( F_D \), linearly, and both forces followed the same trajectory upon decreasing loads, even if the load was increased past the point of damage.

3. DISCUSSION

Mussel tenacity in the high-energy intertidal zone depends critically on and is defined by the vagaries of friction. Friction mediates plaque adhesion but also leads to abrasive damage to
the byssus and catastrophic failure. Mussel byssus attachment tenacity in the high-energy intertidal zone depends critically on friction; we found that damage mitigation can also result in abrasive damage to the byssus and catastrophic failure. The mussel byssus has adapted a hard cuticular coating to resist frictional damage from sand particles and debris. However, having a hard cuticle to confer wear resistance raises concerns about other damage modes such as frictional damage within the cuticle between the softer cuticle matrix and harder cuticle granules (Figure 1b).

In this research, we explored frictional damage mitigation by shear- ing mfp-1/HA complex coacervates between two mica surfaces. Cuticle matrix-inspired coacervates were composed of HA and one of three mfp-1 variants: (1) M. californianus mcfp-1, (2) rmfp-1, and (3) rmfp-1-Dopa. We designed mfp-1/HA matrix-inspired blends to be injectable and deliverable in a concentrated form underwater to coat and protect surfaces. mcpf-1/HA coacervates exhibited excellent wear protection on mica with no damage from applied loads, F_D, as high as 300 mN (pressure, P > 2 MPa). Recombinant proteins only partially recapitulated the wear protective effect: surfaces with rmfp-1/HA coacervates were damaged under 10-fold lower loads than surfaces with mcfp-1/HA coacervates. Coacervate surface wear protection was independent of the coefficient of friction but dependent on Dopa and polycation to polyanion mixing ratio. The wear protection in mcfp-1/HA coacervates suggests that retention of a fluidic component in the cuticular matrix may substantially contribute to damage mitigation in the mussel byssus. We began by investigating the conditions under which mcfp-1/HA coacervates. We found that the degree of coacervation was dependent on salt concentration, pH, and polyanion/polycation mixing ratio. When combined, mcfp-1 and HA coacervated over a 10-fold range of salt concentrations, over the tested pH range of 3.3–4.5, and across all tested polyelectrolyte mixing ratios. Recombinant rmfp-1, both with and without Dopa, also coacervated with HA but were more easily destabilized by salt than the mcfp-1/HA coacervates (Figure 2b). The greater salt tolerance of mcfp-1/HA coacervates may be due to the difference in molecular weight between the native and recombinant proteins (MWmcfp-1 = 92KDa vs MWrmfp-1 = 14KDa). Nonetheless, both mcfp-1 and rmfp-1 coacervate systems delivered concentrated material underwater. To our knowledge, this is the first report of a complex coacervate made from native mussel foot proteins.

Widespread use of complex coacervates in coating and encapsulation technologies for granules, drugs, and perfumes depends critically on their adhesion/cohesion energy, E_ad, and interfacial energy, γ—if. We investigated the adhesion/cohesion energy and the interfacial energy to determine how thoroughly mfp-1/HA coacervates coat surfaces. Coacervates were injected as microdroplets into the SFA and allowed to settle on and coat a mica surface. All tested mfp-1/HA coacervates adhered to mica (0.42 ± 0.12 < E_ad < 1.08 ± 0.25 mJ/m²) and exhibited low interfacial energy (0.28 ± 0.08 < γ—if < 0.72 ± 0.17 mJ/m²) (Figure 5). mcfp-1/HA coacervate at a 1:1 mixing ratio had the lowest interfacial energy (γ—if = 0.28 ± 0.08 mJ/m²) of all the coacervate systems tested, exhibiting the lowest γ—if ever reported for liquid–liquid separations. Visual investigations confirmed that all tested coacervates efficiently coated the surfaces (Figure 4), making them well suited for coating cuticle granules and encapsulation technologies.

After coating the mica surface with the mfp-1/HA coacervate, we investigated frictional damage mitigation by exposing the coacervate-covered mica surface to shear in the SFA. Here, wear protection is defined as the maximum load, F_D, that coacervate-coated mica surfaces withstood before exhibiting shear-induced damage (Figure 4). mcfp-1/HA coacervate mitigated frictional damage to mica surfaces, even under applied normal loads of 300 mN (pressure, P > 2 MPa). rmfp-1/HA coacervate achieved only a fraction of the wear protective effect; shear induced damage with rmfp-1/HA coacervate under 10-fold lower loads than with mcfp-1/HA coacervate. That coacervated native protein dissipated energy and resisted frictional wear to surfaces suggest that the cuticle matrix plays a crucial role in mitigating shear-induced damage in the byssus (Figure 1).

We did not expect this damage mitigation because our coacervates exhibited a comparatively high coefficient of friction, μ = 0.3 (rmfp-151/HA coacervate, μ = 1.3) and synovial fluid, μ = 0.15. However, the coefficient of friction has increasingly been shown to be an invalid measure of surface damage: several studies of lubricants with a low coefficient of friction report poor wear protection to mica surfaces and especially to biological tissues. Our observation that mfp-1/HA coacervates with the same coefficient of friction (μ = 0.3) vary more than 10-fold in damage mitigation supports the notion that damage mitigation is dependent on more than just the coefficient of friction; we found that damage mitigation correlates to both polyanion-to-polycation mixing ratio and surface adhesion. Mfps adhere readily to mica surfaces through a combination of bidentate hydrogen bonding of Dopa and electrostatic interactions between positively charged lysine and negatively charged mica. The recombiant protein lacks Dopa that can be added to the protein to demonstrate the effect of surface adhesion to wear protection. In the absence of Dopa, rmfp-1/HA coacervate is unable to remain attached to mica, allowing frictional stresses to act directly on the mica surface, resulting in...
surface damage under low loads ($F_D < 10 \text{ mN}$, Figures 6 and 7). Addition of Dopa in rmfp-1 increased surface adhesion and protected the surfaces from damage under 5-fold higher loads ($F_D < 60 \text{ mN}$) (Figures 6 and 7). We propose that under compressive shear, mfp-1/HA coacervates with Dopa (rmfp-1-Dopa/HA and mcfp-1/HA) undergo redistribution where Dopa mediates adhesion between mfp-1 and mica. In this way, the surfaces are protected from damage by shifting the slip-plane up from the surface into the intervening coacervate layer, a reconfiguration that has been described as incompressible “molecular ball bearings.”

To further investigate the mcfp-1/HA coacervates, we adjusted the polycation-to-polyanion mixing ratio (7:3 and 3:7) and found that surface wear protection relied on the careful coordination of the mixing ratio. Although mcfp-1/HA coacervates at a 1:1 mixing ratio protected the surfaces from damage at high loads, mcfp-1/HA coacervated at suboptimal 7:3 and 3:7 mixing ratios weakly protected surfaces, which were damaged under only a quarter of the applied load (Figures 4a and 5). In this instance, damage was not due to insufficient adhesion; mcfp-1/HA coacervates with an excess of mcfp-1 or HA had higher adhesion/cohesions energies. Perhaps these molecules not involved in balanced electrostatic interactions (coacervate formation) become entangled, allowing shear forces to impact and damage the surface. The observation that as adhesion increases, wear protection also increases up to a point suggests that adhesion has an optimum for mitigating damage.

Mussel byssal cuticle toughness and durability is widely attributed to covalent cross-links and Ca$^{2+}$ and Fe$^{3+}$ complexation. However, our results raise the interesting possibility that maintaining coacervates (fluid components), perhaps stabilized by microenvironments in the cuticle matrix, could be adaptively beneficial for damage control as well as self-healing since fluids spontaneously self-heal. The excellent frictional damage mitigation by mcfp-1/HA coacervates reveals a significant untapped potential for coacervates in applications that require both adhesion and energy-dissipative lubrication, such as artificial joints, contact lenses, dental sealants, and hair and skin conditioners.

**Figure 7.** Schematic of the proposed wear protection mechanism. Coacervates, condensed by numerous transient electrostatic interactions between lysine and carboxylate moieties (purple asterisk), (a) isotropically distribute in the bulk solution and (b) distribute asymmetrically under compressive shear with mfp-1 (yellow) preferentiality bound to the surface. Adding Dopa to mfp-1 increases adhesion to mica (red asterisk) and increases wear protection by shifting the slip-plane away from the mica and into the intervening layer enriched with HA (green).

**4. CONCLUSIONS**

We delivered mussel cuticle matrix-inspired blends in a concentrated form underwater as complex coacervates. They coated surfaces, dissipated energy, and protected surfaces from wear. Damage mitigation was independent of the coefficient of friction and dependent on surface adhesion and coacervation conditions. On the basis of these results, we propose that during compressive shear, coacervated mfp-1 and HA undergo redistribution whereby Dopa mediates adhesion between mfp-1 and mica. The adhesive layer protects the mica surface by shifting the slip-plane up from the surface into the intervening coacervate layer. mcfp-1/HA coacervates exhibited exceptional surface damage mitigation against shear-induced wear, suggesting that retention of a fluidic component in the cuticular matrix may substantially contribute to damage mitigation in the mussel byssus.

**5. MATERIALS AND METHODS**

**5.1. Purification of Native mcfp-1.** We purified native mcfp-1 as described previously, with some modifications. We harvested mussels from the pilings of Goleta Pier (Santa Barbara, CA) and held them in circulation tanks. The mussels were shucked, and their feet were amputated and stored at $-70^\circ \text{C}$ before freezing off the pigmented epithelium. Approximately 50 prepared feet were homogenized in four equivalents (w/v) of 5% acetic acid (v/v), 10 μM leupeptin, 10 μM pepstatin, and 1 mM ethylenediaminetetraacetic acid in a glass Kontes tissue grinder (Vineland, NJ) on ice, then centrifuged at 20,000g and 4 °C for 40 min. The supernatant was acidified to 1.5% (v/v) perchloric acid. After centrifugation at 20,000g and 4 °C for 40 min, the supernatant was dialyzed for 4 rounds in 4 L of 5% acetic acid (v/v) for 4 h (1000 Da molecular weight cutoff tubing from Spectrum Industries, Los Angeles, CA) before lyophilization. Lyophilized protein was resuspended in 200 μL of 5% acetic acid (v/v), and 50 μL aliquots were run through a Shodex KW-803 size exclusion column (5 μm, 8 × 300 mm) (Showa Denko, New York, NY). Fractions were monitored at 280 nm, and those that absorbed were assessed for sample purity by acid-urea polyacrylamide gel electrophoresis [7.5% acrylamide, 0.2% N,N-methylenebis-acrylamide], 5% acetic acid, and 8 M urea gels then stained with Coomassie Blue R-250 (Sigma-Aldrich, Brooklyn, NY) and amino acid analysis. Pure mcfp-1 fractions were pooled and aliquoted before lyophilization and stored at $-70^\circ \text{C}$ for future use.

**5.2. Recombinant mfp-1 (rmfp-1) and Enzymatic Modification of mfp-1 Tyrosine to Dopa.** rmfp-1 was prepared as in Zheng...
et al.\textsuperscript{41} and was provided by D.S. Hwang (Pohang University of Science and Technology University, Pohang, S. Korea). mfp-1 consisted of the sequence [AKPSYPPYTK]\textsubscript{12} (with a point mutation Ser65 to Cys65). The decapetide repeat sequence of mfp-1 is the same as that of the \textit{M. californianus} decapetide repeat sequence except that the first three positions in the recombinant repeat sequence (AKP\textsuperscript{*}) correspond to the repetitive sequence found in \textit{M. galloprovincialis}, which varies slightly from that of the \textit{M. californianus} repeat (P\textsuperscript{*}KI). Proteins having extensive internal repeat sequences are not well tolerated by & strength, pH, and mixing ratio. 

Amino acid analysis of the final product indicated a Tyr to Dopa conversion of \textasciitilde83\% in mfp-1 (further referred to as mfp-1-Dopa). 

5.3. Coacervation of mfp-1 and HA: Effects of Ionic Strength, pH, and Mixing Ratio. We quantified coacervate yield under different solution conditions via turbidimetric measurements. Each solution was prepared by dissolving either mfp-1 or HA (0.3 mg/mL in sodium acetate buffer [10 mM at pH 3.7 or 4.7 and either 10 mM NaCl (mfp-1 and mfp-1-Dopa) or 60 mM NaCl (mcfp-1)]). Because maximum coacervation occurs with polymers of matching molecular weights,\textsuperscript{43} mfp-1 (14 kDa) was matched with a 35 kDa version of HA, and mcfp-1 (92 kDa) was matched with a 76 kDa HA (LifeBiomedical, Chaska, MN). 

For mixing ratio measurements, we added HA (0.3 mg/mL) to a mfp-1 solution (0.3 mg/mL) at varying volume ratios in sodium acetate/acetic acid buffer (pH 3.7). We investigated ionic strength dependence by adjusting the NaCl concentration at a 1:1 weight ratio for both mcfp-1/HA and mfp-1/HA coacervates using Lys pK\textsubscript{a} = 10.5 and HA-COOH pK\textsubscript{a} = 2.9\textsuperscript{48} in sodium acetate/ acetic acid buffer (pH 3.7). 

5.4. Microscopic Confirmation of the Coacervate Phase. We mixed mfp-1 and HA just prior to their injection (<1 min) between a glass slide and coverslip separated by double-sided tape. Light microscope images were taken with a Zeiss Axioplan microscope equipped with an epiplanochrom microscope camera at 100x magnification 10 min postinjection to monitor coacervate formation. 

5.5. Measuring Normal and Lateral Interactions. We measured the normal and lateral forces as a function of the separation distance, \( D \), between surfaces using the SFA as before.\textsuperscript{45,46} with some modifications. Mica was used because it provides a sensitive testing ground for frictionally induced damage as it is unable to endure shear and then gluing a thin, freshly cleaved mica sheet (1 \( \mu \)m) onto a glass slide and coverslip separated by double-sided tape. Light microscope images were taken with a Zeiss Axioplan microscope equipped with an epiplanochrom microscope camera at 100x magnification 10 min postinjection to monitor coacervate formation. 

We mixed mfp-1 (0.3 mg/mL) and HA (0.3 mg/mL) less than 1 min prior to injecting 50–100 \( \mu \)L of the mixture between the two mica surfaces in the SFA where the mfp-1/HA coacervate coalesced, settled on, and coated the mica surface (30 min, Figure 4). Alternatively, coacervates may be premixed and used in their dense phase; however, this increases time requirements and results in material loss. The force of adhesion (\( F_{\text{ad}} \)) was used to calculate the interfacial energy (\( \gamma_{\text{mfp-1/HA}} = F_{\text{ad}}/3\pi R \)) [the energy required for a coacervate and create new, solvent (equilibrium phase) exposed surface]\textsuperscript{33} and the interaction (adhesion/cohesion) energy by the Derjaguin approximation, \( W_{\text{ad}} = F_{\text{ad}}/2\pi R \). 

We sheared the surfaces at a sliding velocity of 10–100 \( \mu \)m/s with a frictional device.\textsuperscript{46} The coefficient of friction (\( \mu \)) is defined as the slope of \( F_{\text{f}} \) versus \( F_{\text{f}} \) plot. Abrasions in the FECO and Newton rings indicated damage to the mica surface (Figure 4). The SFA allows for verification of a continuous coacervate coating. For every experiment, we confirmed a continuous coacervate coating of the surface through continuity of the FECO and measured the normal and lateral forces without sample disruption. All SFA experiments were performed at room temperature (thermostated at 23 °C).
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